

# siRNA Therapeutics: Target Identification, Discovery and Early Development Considerations

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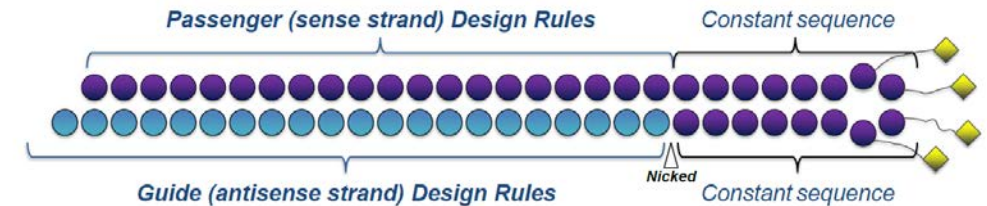
Senior Scientist

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Disclosure: I am an employee of Arrowhead Pharmaceuticals

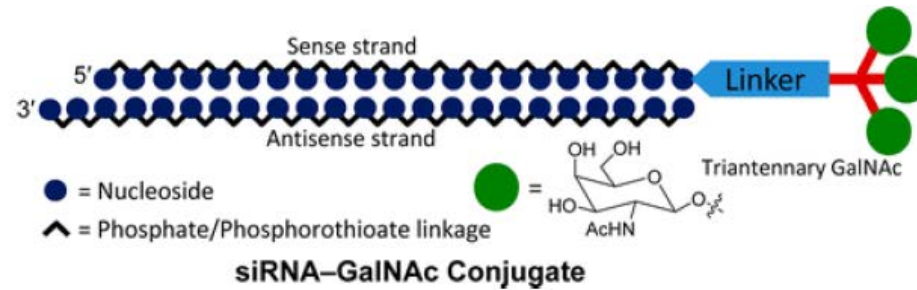
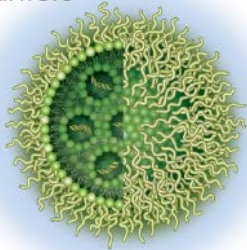
# Multiple Older Platforms converging on targeting with direct conjugation of NAG

- Dicerna – GalXC™ (tetraloop)

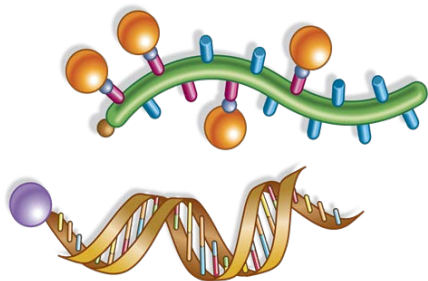


- Alnylam – Lipid Nanoparticle (LNP) to GalNAc conjugation (GalNAc-ESC/ESC+)

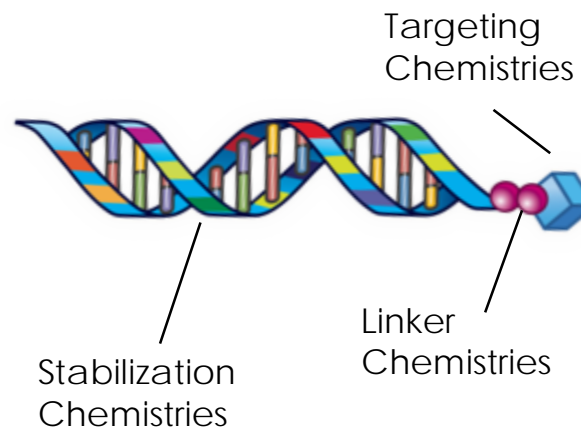
Lipid Nanoparticle



- Arrowhead – Dynamic Polyconjugates (DPC™) (2 molecules with endosomal escape) to TRiM™



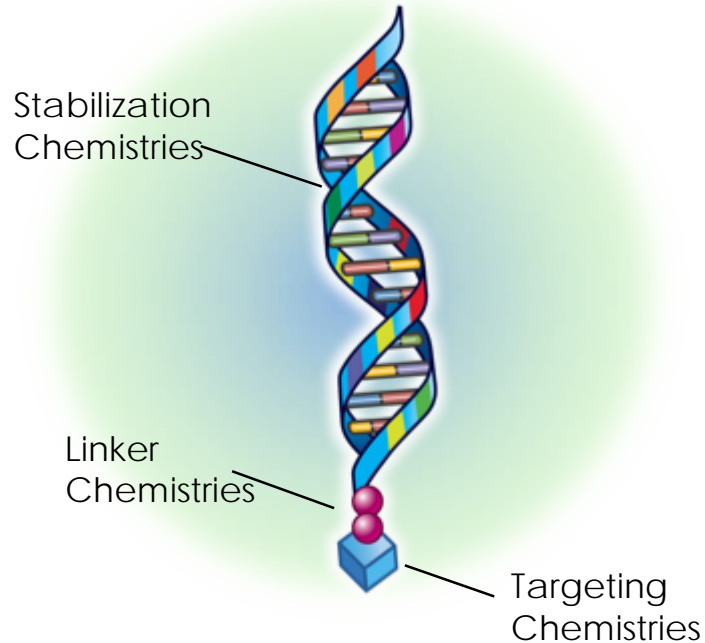
**DPC™ (EX-1) and cholesterol-lined RNAi trigger they are separately targeted to the liver**



**Targeted RNAi Molecule TRiM™ platform**

# Arrowhead RNAi Platform: TRiM™

## Simplicity, Specificity, and Activity



TRiM™ has rules and algorithms to optimize trigger sequence

- Limit cross reactivity with off target genes
- Maximize activity
- Maximize innate stability
- Rational use and placement of modifying chemistries
- RNAi chemistry insights and expertise have allowed us to see what others have not

Targeted RNAi Molecule  
TRiM™ platform

# Hepatic siRNA Discovery/Development

Direct conjugation with NAG allows for binding and endocytosis with highly and specifically expressed Asialoglycoprotein receptor (ASGPr) in hepatocytes

Binding of NAG to ASGPr initiates endocytosis

## Key Design Elements in Hepatic Platform

- Subcutaneous dosing, monthly or less dosing frequency
- Stable and potent sequences
  - No need for the use of endosome escape moieties
- Suppression of liver production of target protein
- Expectation of wide therapeutic index

# What makes an optimal hepatic RNAi gene target

Examine diseases with limited or no treatment options, where knockdown of protein expression is hypothesized to be beneficial to disease initiation/progression

With Hepatocyte-targeted RNAi agents:

- Target is expressed in hepatocytes
  - If not primarily expressed in hepatocytes, hepatocyte expression is key for disease etiology
- Because of ease and familiarity for markets/regulators, we prefer targets that are not easily/well targeted with small molecules or mAbs
- Advantageous if sequence is cross-reactive with human, NHP, and rodent
- Secreted protein advantageous (blood-based monitoring of knockdown)
- Non-secreted protein knockdown can be monitored through liver biopsy or well characterized secondary biomarker
- Disease-relevant animal models available
  - Proof of Concept studies
  - Can be used to estimate level of knockdown required for beneficial effect

2 examples of targets: Factor XII (F12) for thrombotic disease and Alpha 1 antitrypsin (AAT) for AAT-deficiency

# Hepatic RNAi agent development funnel

Bioinformatic selection of RNAi trigger sequences specific for target gene – filter to identify cross-reactive triggers (human/NHP/rodent, human/NHP)



Cross-reactive RNAi trigger synthesis and *in vitro* testing



Synthesis and *in vivo* testing of select RNAi triggers amenable for subcutaneous (SC) administration



Lead Optimization on RNAi triggers for SC administration with *in vivo* testing



Proof of Concept for disease modification in animal models

Exploratory Toxicology

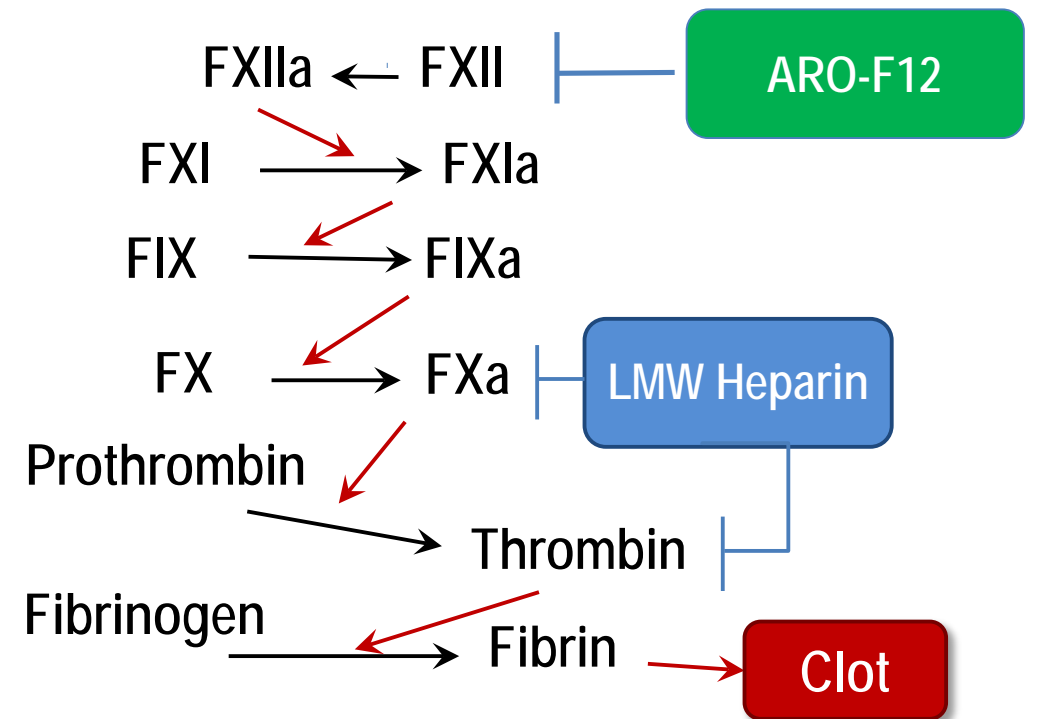
# Targeting Factor XII by RNAi as a prophylactic treatment of thrombotic disease

## Factor XII (F12)

- Key component of contact activation pathway (thrombosis) and kinin-kallikrein (angioedema)
- Predominantly expressed in the liver; circulates in plasma

## F12 inhibition is genetically validated

- F12-deficient mice:
  - viable and fertile<sup>4</sup>
  - do not show bleeding defects<sup>4,5</sup>
  - protected from thromboembolic disease (stroke, pulmonary embolism)<sup>5</sup>
- F12 deficiency in humans is not associated with either bleeding or thrombotic disorders<sup>1,2,3</sup>



<sup>1</sup> Girolami A. *et al.* (2004) *J. Thromb. Thrombolysis* 17:139–143

<sup>2</sup> Koster A. *et al.* (1994) *Br. J. Haematol.* 87:422–424

<sup>3</sup> Zeerleder S. *et al.* (1999) *Thromb. Haemost.* 82:1240–1246

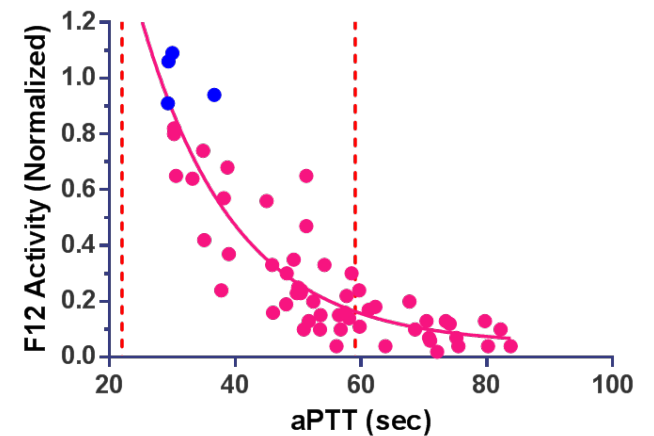
<sup>4</sup> Pauer, H. U., *et al.* (2004) *Thromb. Haemost.* 92:503

<sup>5</sup> Renne, T. *et al.* (2005) *J. Exp. Med.* 202:271

\* Figure modified from Albert-Weissenberger, C., *et al.* (2014) *Front. Cell Neurosci.* 8:345

# Measuring F12 knockdown and effects – Serum/Plasma

- F12 levels can be measured in mouse and NHP by ELISA-based methods to monitor knockdown
  - RNAi triggers tested are cross reactive between human, NHP and rodent
  - Mouse F12 protein (total and activated) measured by custom AlphaLISA™ (Perkin Elmer)
  - NHP F12 protein measured by human F12 ELISA (cross-reactive with NHP)
- F12 activity can be measured through a modified version of standard coagulation measure activated Partial Thromboplastin Time (aPTT)
- aPTT is inversely correlated with F12 levels (ELISA or Activity)

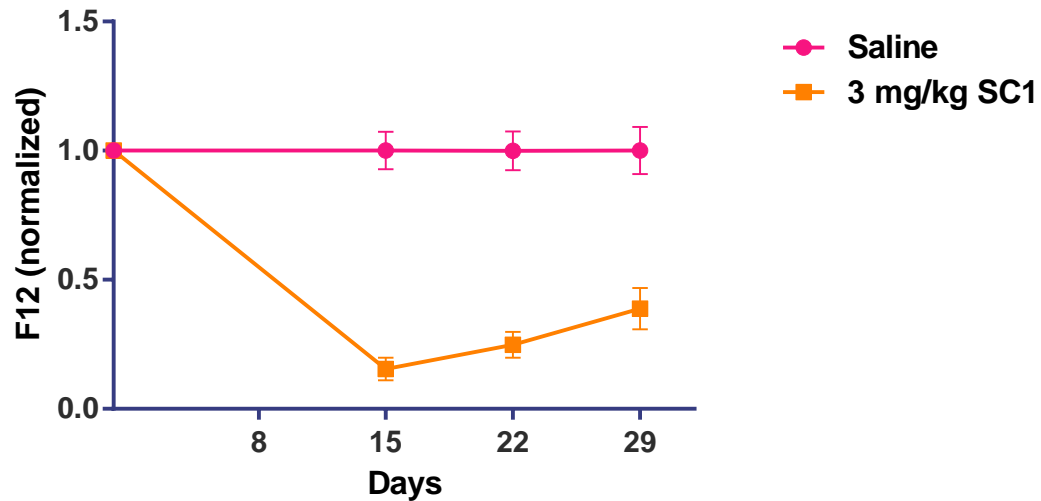




# Examination of modified RNAi triggers in mice

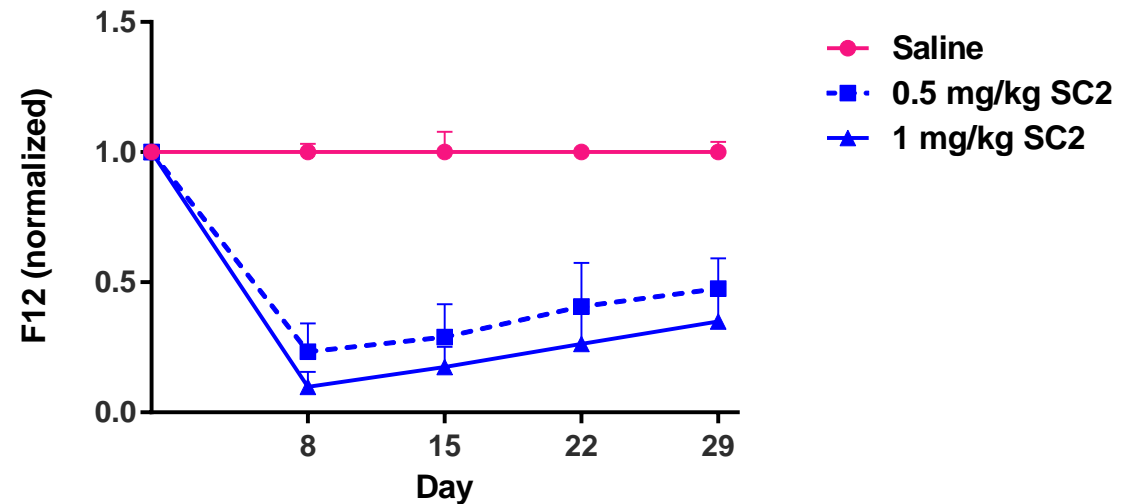
## First Generation

Single 3 mg/kg SC dose  
n=3/group



## Second Generation

Single 0.5 or 1 mg/kg SC dose  
n=4/group

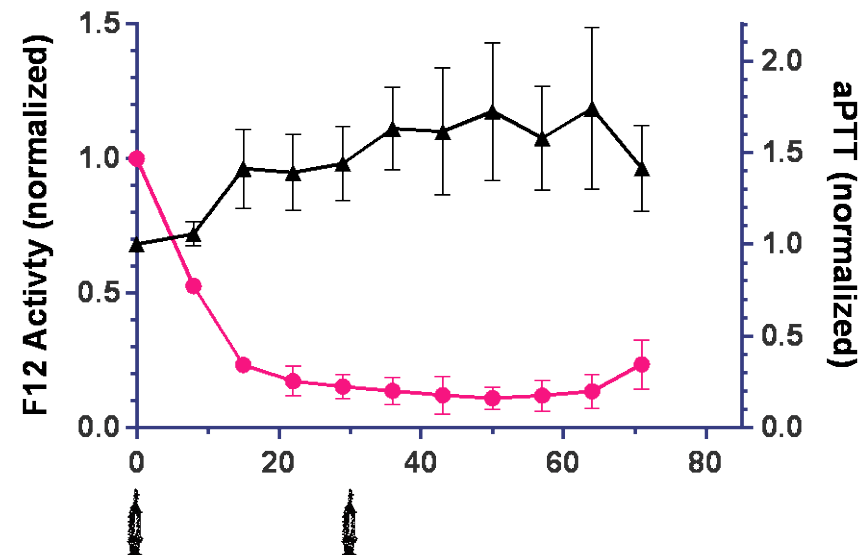
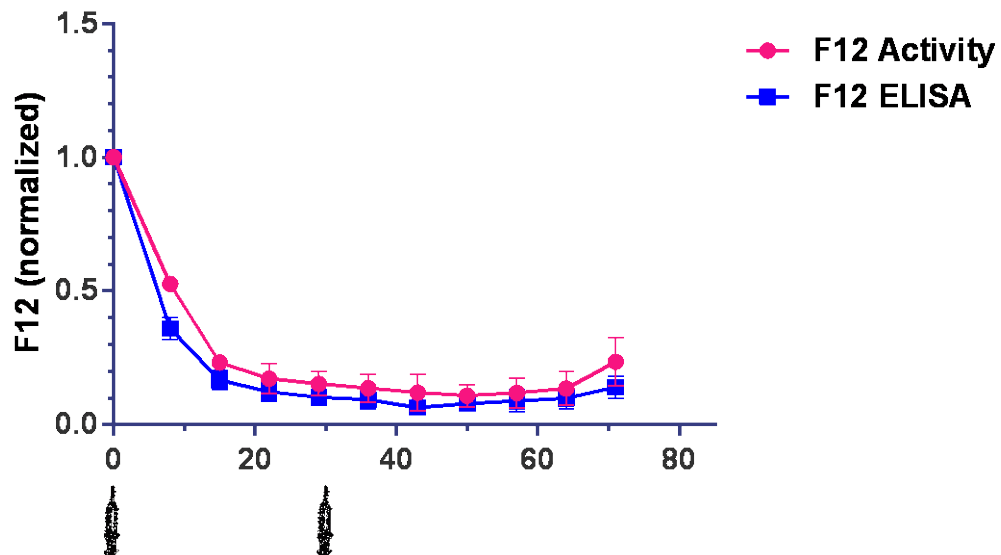


- Modifications to SC1 to yield SC2 improved knockdown
  - 85% at 3 mg/kg vs 91% at 1 mg/kg at nadir
- Dose response observed with SC2

# Second Generation Triggers – Examination in NHP

- Initial SC dose of 3 mg/kg SC2, followed by 1.5 mg/kg dose on day 29

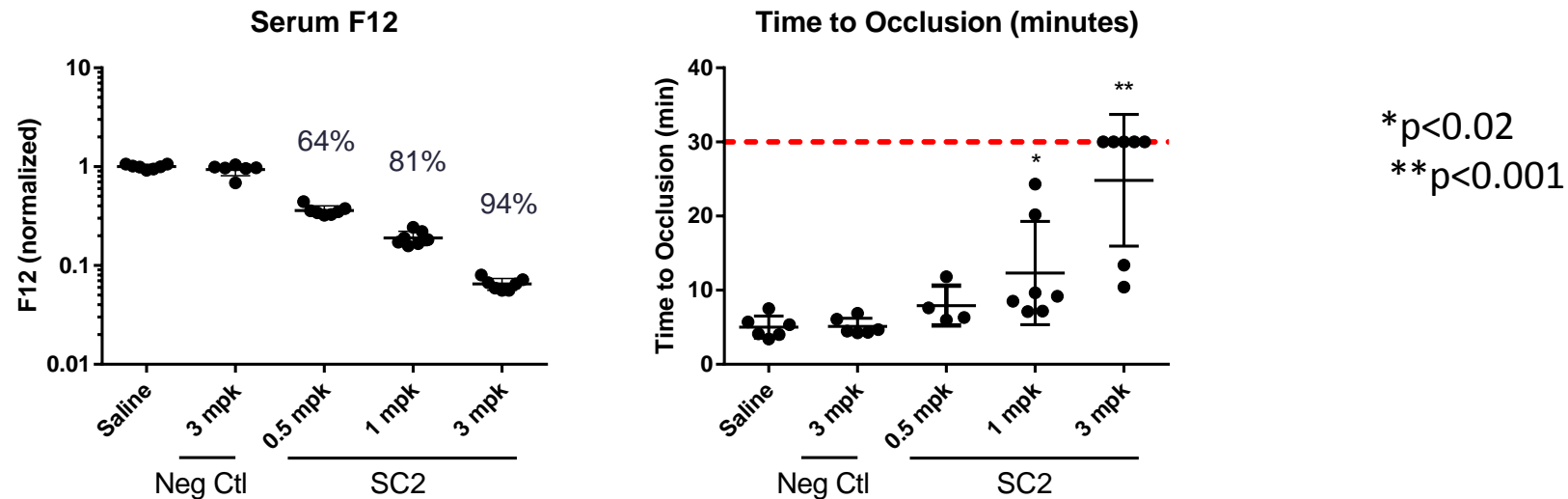
*n=2/group*



- Achieved ~90% knockdown of F12 in NHP after the second dose at 1.5 mg/kg with >1 month duration
- 90% knockdown of F12 activity correlates with significant increase in aPTT
- No changes in toxicity markers (clin chem, CBC) after dosing

# Disease-relevant Animal Modeling: Ferric-chloride study

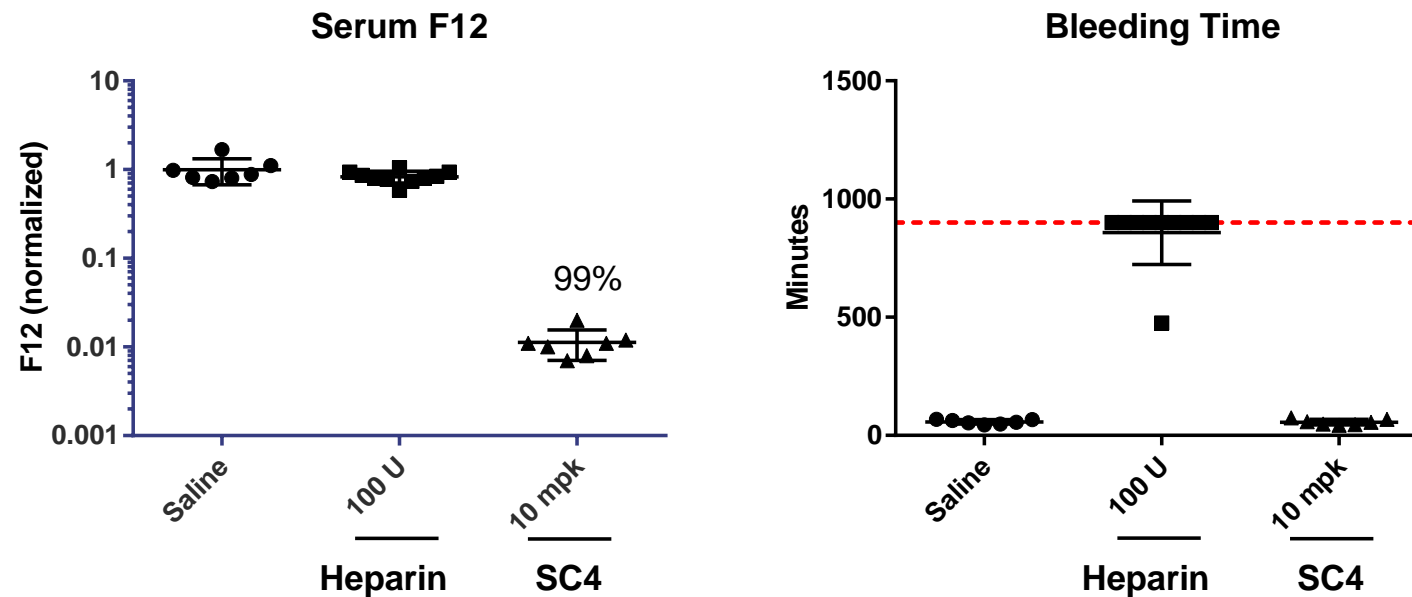
- *Thrombus induced by exposure of carotid artery to  $FeCl_3$*
- *Measure time to blood flow occlusion (thrombus formation)*
- *Single SC injection of SC2 or negative control, 2 weeks prior to challenge with  $FeCl_3$ ,  $n=7/group$*



- Dose response observed for inhibition of clot formation
- Statistically significant change in occlusion times ( $p<0.02$ ) observed with  $>80\%$  knockdown of serum F12

# Bleeding risk assessment through mouse modeling

- *Transverse cut of tail vein, monitor time to clotting*
- *Single dose SC4, 14 days prior to assessment, n=7/group (saline and SC4), n=10/group (heparin)*



- No increased bleeding observed, even with 99% knockdown of F12 levels
- Consistent with F12 <sup>(-/-)</sup> mice showing no increase in bleeding over wild type controls

# Alpha-1 Antitrypsin Deficiency (AATD)

- AATD is a large scale orphan disease
  - Alpha-1 Foundation estimates 100,000+ in the US
  - Approximately 100,000+ in Europe
- Mutation in AAT gene (Z-AAT) leads to mis-folding of the protein and poor export from hepatocytes: low levels in circulation and accumulation in liver

## Pathophysiology

### Lung

Tissues susceptible to damage by neutrophil proteases: COPD



Treated with AAT enzyme replacement therapy

### Liver

Accumulation of mutant Z-AAT protein can cause cirrhosis and HCC



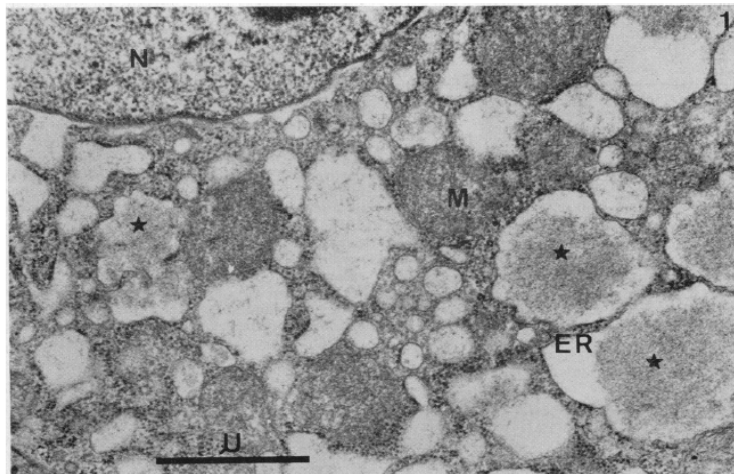
**Currently no treatment**

# RNAi trigger mechanism of action

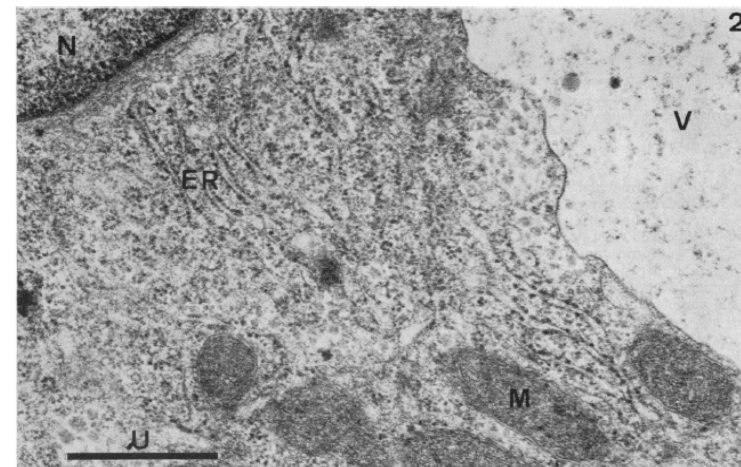
RNAi trigger designed to stop Z-AAT production by silencing AAT gene to:

- Prevent liver accumulation
- Allow clearance of accumulated protein
- Prevent cycles of cellular damage
- Prevent/Reverse progression of liver fibrosis

**PiZZ phenotype (diseased)**

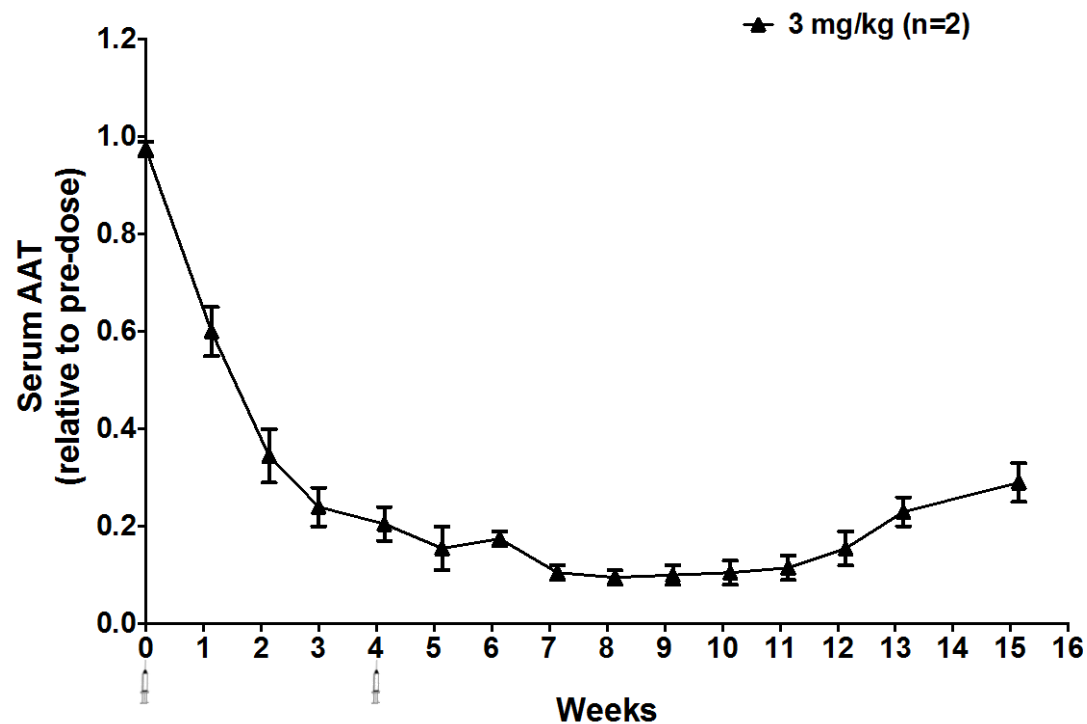


**Pi null phenotype (normal)**



# ARO-AAT Provides Durable AAT knockdown: Multi-dose in NHP, dosed subcutaneously

- 92% maximum serum AAT knockdown achieved in cynomolgus monkeys
- Knockdown sustained for 7+ weeks following second dose



Durable knockdown supports once monthly or less frequent dosing

# Exploratory Toxicology: ARO-AAT preliminary safety evaluation

- Based on clinical observations, clinical pathology and limited histopathology evaluations, ARO-AAT was well tolerated in the following non-GLP exploratory toxicity studies:
  - A repeated dose study in rats administered 3 weekly subcutaneous doses at dose levels of 30, 60, 120, and 300 mg/kg
  - An escalating dose study in two cynomolgus monkeys dosed weekly subcutaneously at doses up to and including 300 mg/kg



# Key Considerations entering development

- FDA treats RNAi therapeutics like small molecules (CDER)
- Requirements for particular enabling studies may vary based on placement within CDER
- Coordination of required GLP studies can speed transition to clinic

# Summary

- Most current RNAi agents specifically target hepatocytes through direct conjugation with NAG (ASGPr1 ligand)
- RNAi agents can be effective in knocking down expression of target protein responsible for rare/orphan diseases (AAT-deficiency) and more common conditions (Factor XII in thrombosis)
- Speed of evaluation of potency/efficacy is increased with human/NHP/rodent cross-reactive RNAi agents
- Knockdown that can be measured by blood biomarker (primary or secondary biomarker) speeds evaluation
  - Non-secreted target protein can be measured by liver biopsy, or other methods
  - ARO-F12 and ARO-AAT displayed durable knockdown of target protein in NHP
- Exploratory toxicology studies of RNAi agents support wide therapeutic index
- RNAi agents are considered as small molecules by regulatory agencies, with respect to requirements

# Arrowhead Team

